

中文題目：在人類支氣管上皮細胞中，缺氧誘導因子 HIF1 $\alpha$  與叉頭框轉錄因子 FOXA2 共定位於細胞核內並下調 FOXA2 所調控的神經內分泌多肽 $\alpha$ -和 $\beta$ -CGRP 基因及緊密連接蛋白基因 *TJP1* 和 *TJP2* 的表現，然而卻顯著上調第二型發炎細胞激素 *IL5*、*IL13* 和 *IL33* 的表現

英文題目：Hypoxia-Inducible Factor HIF1 $\alpha$  Co-localizes with the Forkhead Box Transcription Factor FOXA2 and Down-regulates FOXA2-Mediated Expression of Neuroendocrine Peptide  $\alpha$ - and  $\beta$ -CGRP Genes and the Tight Junction Protein Genes *TJP1* and *TJP2*, whereas Up-regulates the Type 2 Inflammatory Cytokines *IL5*, *IL13* and *IL33* in Human Bronchial Epithelial Cells

作者：李育銘<sup>1</sup>，林昭如<sup>2</sup>，石宇軒<sup>3,\*</sup>，陳怡濤<sup>2,\*</sup>

服務單位：<sup>1</sup>台中榮民總醫院內科部，<sup>2</sup>國防醫學院航太及海底醫學研究所，<sup>3</sup>台中榮民總醫院內科部血液腫瘤科 \*共同通訊作者

**Background:** Our recent study has reported that human bronchial epithelial cells (HBECs) cultured under both consecutive hypoxia and intermittent hypoxia-reoxygenation (H/R) display significantly increased expression of the mucin protein and gene MUC5AC, which is associated with increased expression of the hypoxia-inducible factor HIF1 $\alpha$  under both hypoxia and H/R. Notably, previous studies have shown that MUC5AC expression is up-regulated by Th2 inflammatory cytokines including IL-5 and IL-13, and down-regulated by the forkhead box transcription factor FOXA2. To further decipher the molecular mechanisms underlying the regulatory interplay between hypoxia, HIF1 $\alpha$ , FOXA2, and Th2 inflammatory cytokines in human airway epithelium, it is of interest to study the effects of consecutive hypoxia, intermittent H/R, and changes of HIF1 $\alpha$  expression levels on the expression of Th2 inflammatory cytokines, FOXA2, and the downstream target genes of FOXA2, including the neuroendocrine cell marker calcitonin gene-related peptides  $\alpha$ - and  $\beta$ -CGRP (encoded by *CALCA* and *CALCB* genes, respectively), and the epithelial barrier tight junction proteins ZO-1 and ZO-2 (encoded by *TJP1* and *TJP2* genes, respectively).

**Method:** The normal (NHBECS) and COPD-diseased (DHBECS) human bronchial epithelial cells were each derived from three distinct age-matched Caucasian donors and were all obtained from the Lonza Biotechnology Company in the U.S.A.. The HBECs were either transfected with a scrambled siRNA or empty cDNA vector, or transfected with *HIF1A* siRNA or *HIF1A*-overexpressing cDNA vector, followed by air-liquid interface (ALI) culturing under normoxia (21% O<sub>2</sub>) for 3 days for cell proliferation, and subsequently cultured in the differentiation medium consecutively under 21% O<sub>2</sub> for another 18 days, or under 24/24-hour cycles of intermittent hypoxia-reoxygenation (H/R) (i.e., 1% O<sub>2</sub> and 21% O<sub>2</sub> alternately) for 18 days in total, or cultured consecutively under 1% O<sub>2</sub> for 9 days in total, followed by returning to 21% O<sub>2</sub> for another 9 days in total. Total mRNAs were then extracted from NHBECS and DHBECS cultured under different oxygen tensions, followed by microarray analyses, qPCR analyses and immunofluorescence staining.

**Results:** Both consecutive hypoxia and intermittent H/R significantly down-regulated expression of both  $\alpha$ - and  $\beta$ -CGRP genes *CALCA* and *CALCB* as well as their upstream activators, the transient

receptor potential vanilloid type 1 gene *TRPV1* and the forkhead box gene *FOXA2*, and also the tight junction protein genes *TJP1* and *TJP2*, whereas up-regulated the type 2 inflammatory cytokine genes *IL5*, *IL13* and *IL33*, in both NHBEs and DHBECs. Transfection with *HIF1A* siRNA into HBECs at the beginning of the ALI culturing was sufficient to up-regulate *CALCA*, *CALCB*, *TRPV1*, *TJP1* and *TJP2* mRNA expression to the levels comparable with the expression levels under normoxia, while down-regulating Th2 cytokine genes *IL5*, *IL13* and *IL33* to the expression levels comparable to those under normoxia. Interestingly, transfection with *HIF1A*-overexpressing cDNA vector into HBECs cultured under normoxia was sufficient to significantly decrease the mRNA levels of *CALCA*, *CALCB*, *TRPV1*, *TJP1* and *TJP2* and significantly increase the mRNA levels of *IL5*, *IL13* and *IL33*. Nonetheless, it is noteworthy that neither transfection with *HIF1A* siRNA nor transfection with *HIF1A* cDNA significantly altered the mRNA or protein level of *FOXA2*, indicating that *FOXA2* expression is not subject to regulation by *HIF1α*.

Notably, our immunofluorescence staining analyses indicated that CGRP immunostaining signals were detected in *HIF1α*<sup>-</sup>/*FOXA2*<sup>+</sup> cells only, whereas MUC5AC immunostaining signals were detected in *HIF1α*<sup>+</sup>/*FOXA2*<sup>-</sup> cells only, in agreement with the respectively positive and negative regulation of the CGRP and MUC5AC genes by *FOXA2*, and in contrary the respectively negative and positive regulation of the CGRP and MUC5AC genes by *HIF1α*.

**Conclusion:** Taken together, our study shows for the first time that *HIF1α* in human airway epithelial cells is capable of down-regulating *TRPV1*, *CALCA*, *CALCB*, *TJP1* and *TJP2* mRNA expression, and may regulate gene expression of the Th2 inflammatory cytokines, including the pro-tumorigenic cytokine IL-33, either directly or indirectly via CGRP inhibition. Down-regulation of the tight junction proteins *TJP1* (*ZO-1*) and *TJP2* (*ZO-2*) expression by *HIF1α* antagonizes the anti-EMT (epithelial-to-mesenchymal transition) effect of *FOXA2* and may facilitate tumor metastasis. It remains to be studied whether *HIF1α* and *FOXA2* mutually inhibits the transactivation activity of each other in the airway epithelium.

